
Deep Contextual Learners for Protein Networks

Michelle M. Li¹ Marinka Zitnik^{1 2 3}

Abstract

Spatial context is central to understanding health and disease. Yet reference protein interaction networks lack such contextualization, thereby limiting the study of where protein interactions likely occur in the human body and how they may be altered in disease. Contextualized protein interactions could better characterize genes with disease-specific interactions and elucidate diseases' manifestation in specific cell types. Here, we introduce AWARE, a graph neural message passing approach to inject cellular and tissue context into protein embeddings. AWARE optimizes for a multi-scale embedding space, whose structure reflects network topology at a single-cell resolution. We construct a multi-scale network of the Human Cell Atlas and apply AWARE to learn protein, cell type, and tissue embeddings that uphold cell type and tissue hierarchies. We demonstrate AWARE's utility on the novel task of predicting whether a protein is altered in disease and where that association most likely manifests in the human body. To this end, AWARE outperforms generic embeddings without contextual information by at least 12.5%, showing AWARE's potential to reveal context-dependent roles of proteins in disease.

1. Introduction

Modeling interactions between proteins has been crucial for disentangling the rich diversity of complex biological phenomena, such as the mechanisms underlying organ function in multicellular organisms (Luck et al., 2020; Bassett & Sporns, 2017) and disease processes (Cheng et al., 2018; Menche et al., 2015; Sonawane et al., 2019). However, protein interaction (PPI) networks are typically presented as generic maps without contextual information about the particular cell type, disease state, and external stresses and

stimuli on the system, which are known to have an influence on protein interactions and their function (Przytycka et al., 2010; Ideker & Krogan, 2012; Zhang et al., 2016; Willsey et al., 2018; Markmiller et al., 2018). As a result, there have been many efforts to contextualize protein interactions, which have only further revealed underlying mechanisms of disease progression and drug action (Basha et al., 2020; Greene et al., 2015; Zitnik & Leskovec, 2017). Due to recent advancements in single-cell sequencing technology, we are now better equipped to investigate proteins and their roles in disease across tissues and at a single-cell resolution (Kamies & Martinez-Jimenez, 2020; Saviano et al., 2020).

Integrated single-cell transcriptomics and PPI network maps have already yielded insights into functional modules and gene-interaction dynamics of disease (Klimm et al., 2020; Mohammadi et al., 2019). Prevailing approaches construct cell type specific PPI subnetworks and apply network analytical techniques on them individually (Cha & Lee, 2020). While such contextualized protein networks are powerful resources for descriptive analyses, they cannot generate optimized representations and provide predictions in downstream tasks. They could greatly benefit from the predictive capacity provided by deep graph learning, an emerging area founded upon key principles of network science (Li et al., 2021). A deep graph learning approach could capture networks' topology and generate compact embeddings that could be easily specialized to cellular and tissue contexts.

Here, we develop AWARE, a multi-scale graph neural network (GNN) to learn embeddings at various scales, guided by the hierarchical structure and function of cells and tissues. AWARE integrates cellular and tissue contextual information in the GNN architecture itself. As a result, AWARE can incorporate cell-specific gene expression into PPI networks while injecting the structure of protein interactions and cell type and tissue hierarchies. We evaluate AWARE on the Human Cell Atlas and cell type specific disease-gene associations, and demonstrate that AWARE embeddings outperform generic embeddings by at least 12.5%, thus highlighting the importance of contextual learners for protein networks.

2. Methods

We start by describing the construction of our cell type specific PPI subnetworks, and proceed with an overview of

¹Harvard University ²Broad Institute of MIT and Harvard ³Harvard Data Science. Correspondence to: Michelle M. Li <michelleli@g.harvard.edu>, Marinka Zitnik <marinka@hms.harvard.edu>.

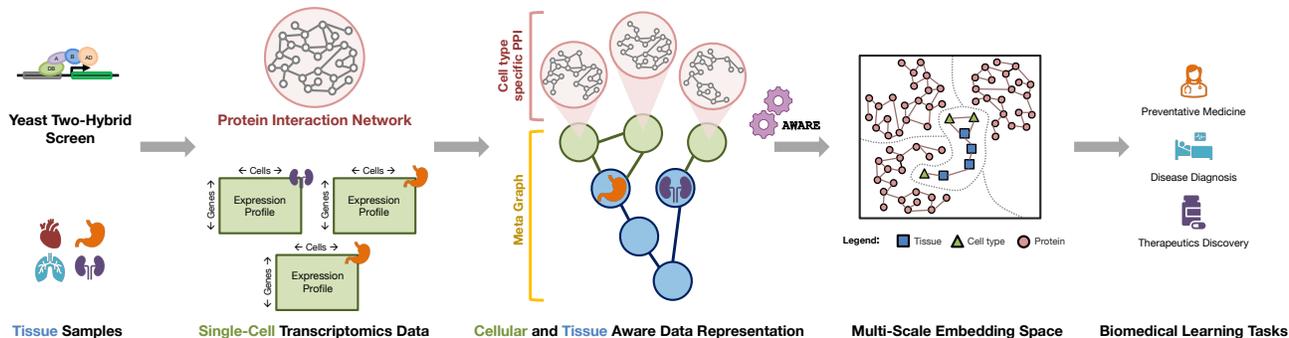


Figure 1. Overview of the approach. We construct cell type specific PPI subnetworks from a global PPI network (predominantly constructed by yeast two-hybrid screens) and single-cell transcriptomics data (usually generated from tissue samples). We inject cellular and tissue context based on single-cell transcriptomics meta-data and the tissue hierarchical ontology to create a data representation on which we apply AWARE to generate multi-scale protein, cell type, and tissue embeddings. Finally, we utilize such embeddings for downstream biomedical learning tasks related to preventative medicine, disease diagnosis, and therapeutics discovery.

our multi-scale graph neural network approach, AWARE.

2.1. Constructing cell type specific protein networks

We extract cell type specific PPI subnetworks from a global PPI network based on differential gene expression without making any modifications. We first process, normalize, cluster, and annotate our single-cell transcriptomics (scRNA-seq) dataset using SCANPY (Wolf et al., 2018) and SCSA (Cao et al., 2020). We then select the top k differentially expressed genes in each cluster (or cell type). We create a subgraph S from these k nodes, and continuously add n nodes until S is connected. Ultimately, each cell type specific PPI subnetwork S_c (for $c \in C$ cell types) consists of the top $k + n$ differentially expressed genes.

2.2. Contextualizing embeddings of protein networks

Our method, AWARE, has four components: (1) *Initializing* cell type specific PPI embeddings using node- and semantic-level attention (Sec. 2.2.1); (2) *Up-pooling* from cell type specific PPI embeddings to update cell type embeddings (Sec. 2.2.2); (3) *Updating* cell type and tissue embeddings using node- and semantic-level attention (Sec. 2.2.1); and (4) *Updating* cell type specific PPI embeddings using node- and semantic-level attention, and *down-pooling* from cell type embeddings to regularize the cell type specific PPI embeddings (Sec. 2.2.2). In the following sections, we explain the details of our node- and semantic-level attention, up- and down-pooling steps, and overall objective function.

2.2.1. NODE- AND SEMANTIC-LEVEL ATTENTION

We aggregate information from neighbors in each meta-path $\phi_r \in \Phi$ (i.e. sequence of node types; see Sun et al. (2011) for the definition of meta-paths) using node- and semantic-level attention mechanisms (Wang et al., 2019).

Our *node-level attention* learns the importance $\alpha_{i,j}^{\phi_r}$ of node n_j to its neighbor n_i connected by meta-path ϕ_r at layer l : $\mathbf{z}_{n_i}^l(\phi_r) = \|\|_{k=1}^K \sigma(\sum_{j \in \mathcal{N}_i^{\phi_r}} \alpha_{i,j}^{\phi_r} \mathbf{M}^{\phi_r} \mathbf{h}_{n_j}^{l-1})$, where K is the number of attention heads, $\|\|$ denotes concatenation, σ is the nonlinear activation function, $\mathcal{N}_i^{\phi_r}$ is the set of neighbors for n_i connected by ϕ_r (includes itself via self-attention), \mathbf{M}^{ϕ_r} is a type-specific transformation matrix to project the features of n_i , and $\mathbf{h}_{n_j}^{l-1}$ is the previous layer’s embedding for n_j . After generating node embeddings for every meta-path, our *semantic-level attention* learns the importance β^{ϕ_r} of each meta-path $\phi_r \in \Phi$ to n_i such that $\mathbf{h}_{n_i}^l = \sum_{\phi_r \in \Phi} \beta^{\phi_r} \mathbf{z}_{n_i}^l(\phi_r)$. We learn protein embeddings via the *protein-protein* meta-path, and cell type and tissue embeddings via the *celltype-celltype*, *celltype-tissue*, and *tissue-tissue* meta-paths.

2.2.2. UP-POOLING AND DOWN-POOLING

Since we define cell types by their most differentially-expressed genes (Sec. 2.1), we up-pool information from the embeddings of such genes in their corresponding PPI subnetworks. We initialize cell type embeddings by taking the average of their proteins’ embeddings, weighted by the proteins’ relative differential expression d_i^c such that $\mathbf{h}_c^0 = \sum_{i \in V_c} d_i^c \mathbf{h}_{n_i^c}^0$, where $\mathbf{h}_{n_i^c}^0$ is the embedding of node $n_i \in V_c$ in the PPI subnetwork for cell type c . We initialize tissue embeddings by taking the average of their neighbors: $\mathbf{h}_t^l = \frac{1}{|N_t^l|} \sum_{i \in N_t^l} \mathbf{h}_i^{l-1}$. For $l > 0$, we up-pool from cell type specific PPI embeddings using an attention mechanism to learn the importance γ_i^c of node n_i to cell type c such that $\mathbf{h}_c^l = \mathbf{h}_c^{l-1} + \|\|_{k=1}^K \sigma(\sum_{i \in V_c} \gamma_i^c \mathbf{h}_{n_i^c}^{l-1})$. Note that we are learning the weights of proteins that are important, which could be useful for identifying novel cell type biomarkers.

After propagating cell type and tissue information in the meta graph via node- and semantic-level attention

(Sec. 2.2.1), we down-pool from cell type c to its corresponding PPI subnetwork by leveraging the learned importance γ_i^c of node n_i to c such that $\mathbf{h}_{n_i}^l = \mathbf{h}_{n_i}^c + \gamma_i^c \mathbf{h}_c^l$. Intuitively, we are imposing the structure of the meta graph onto the PPI subnetworks based on the proteins’ importance to their corresponding cell type’s identity.

2.2.3. OVERALL OBJECTIVE FUNCTION

To capture the structure of cell type specific PPI subnetworks and the meta graph, we apply softmax \mathcal{L}_S on link prediction. Additionally, to optimize meta-graph-based clustering of the cell type specific PPI embeddings, we use center loss (Wen et al., 2016) to predict the associated cell type of each protein: $\mathcal{L}_C = \frac{1}{2} \sum_{i=1}^m \|\mathbf{x}_i - \mathbf{c}_{y_i}\|_2^2$ for m samples. Thus, $\mathcal{L} = \mathcal{L}_S + \lambda \mathcal{L}_C$, where λ is a hyperparameter.

3. Experimental setup and results

Here, we describe our protein, cellular, and tissue datasets, baseline methods, and implementation details.

3.1. Datasets

Global protein interaction network. Our global PPI network is the union of physical multi-validated interactions from BioGRID (Oughtred et al., 2019; Stark et al., 2006), the Human Reference Interactome (Luck et al., 2020), and (Menche et al., 2015) with 15,461 nodes and 207,641 edges.

Human Cell Atlas. We integrate seven 10X scRNA-seq datasets from healthy individuals in the Human Cell Atlas (HCA) (Regev et al., 2017), and obtain 64 cell type specific PPI subnetworks, each with 1,645 proteins on average.

Cell type and tissue hierarchies. The meta graph is composed of cell type and tissue nodes. Cell type nodes are defined by the annotated clusters from scRNA-seq analysis (Sec. 2.1); cell-cell interactions, by CellPhoneDB (Efremova et al., 2020); celltype-tissue relationships, by meta-data from HCA; and tissue hierarchy, by BRENDA Tissue Ontology (Gremse et al., 2010). Here, we use 8 (of 64) cell types and 4 (of 50) tissues from the meta graph (Fig. 2a).

Disease-gene associations at single-cell resolution.

We extract single-cell disease-gene associations from SC2disease (Zhao et al., 2021) for breast, liver, and lung cancers, Alzheimer’s disease (AD), rheumatoid arthritis (RA), autism spectrum disorder (ASD), lupus nephritis, pancreatic ductal adenocarcinoma, multiple sclerosis (MS), and atherosclerosis. Among these diseases, the overlapping cell types are T cells, astrocytes, and monocytes. The global and cell type specific PPI and our SC2disease dataset share 245 genes. We train and evaluate a KNN for predicting cell type specific disease-gene associations on 80% and 20% of the genes, respectively (Tbls. 1-2). In this limited dataset, only

20 labels have enough samples for evaluation.

3.2. Baseline methods and implementation details

To demonstrate the necessity of each component in AWARE, we perform ablation studies on its core features: cell type specificity (Sec. 2.1), meta graph regularization (Sec. 2.2.2), and prototypical loss (Sec. 2.2.3). In GLOBAL, we ignore scRNA-seq information; in -MG, we turn off down-pooling (i.e. remove notion of cellular and tissue hierarchies); in -PROTO, we set $\lambda = 0$ (i.e. do not optimally separate cell type specific protein embeddings). Across all methods, our node feature matrix has dimension 2048; output layer has dimension 128; node- and semantic-level attentions have 8 heads; softmax lr = 0.001 and center loss lr = 0.01. In AWARE, $\lambda = 0.001$. We use PyTorch Geometric, Adam optimizer, and grid-search for hyperparameter tuning.

3.3. Results

Finally, we discuss the results of our experiments to evaluate the quality and utility of AWARE’s embeddings.

Learning cellular and tissue aware embeddings. Unlike GLOBAL and -MG (Fig. 2b), which are unable to discern cell type specificity in the embedding space, the cellular and tissue aware embeddings generated by -PROTO and AWARE (Fig. 2c) cluster according to the meta graph’s structure. While -PROTO has less clear cell-type-separation, it is notable that -PROTO and AWARE can generate highly granular embeddings that capture cell type and tissue hierarchies.

Predicting cell type specific disease-gene associations.

Embeddings generated with some notion of cellular and tissue hierarchy, i.e. AWARE and -PROTO, outperform those without it by at least 12.5% (Tbl. 1). AWARE and -PROTO embeddings have comparable or better performance than GLOBAL embeddings in 15 (of 20) labels (Tbl. 2). In particular, they outperform GLOBAL in predicting genes in T cells associated with breast cancer and MS by up to 50%; in monocytes associated with RA by 200%; and in astrocytes associated with atherosclerosis by 50%. Such improvements highlight the need to contextualize protein networks with biologically meaningful cell type and tissue dependencies.

4. Discussion and conclusion

To inject cellular and tissue context into biological network embeddings, we have developed AWARE, a methodology to generate rich embeddings of proteins, cell types, and tissues that adhere to cell type and tissue hierarchies. We show AWARE’s utility on the novel task of predicting cell type specific disease-gene associations, demonstrating the importance of cellular and tissue context in protein networks. We envision AWARE to open up new possibilities for contextually adaptive embeddings in biomedicine.

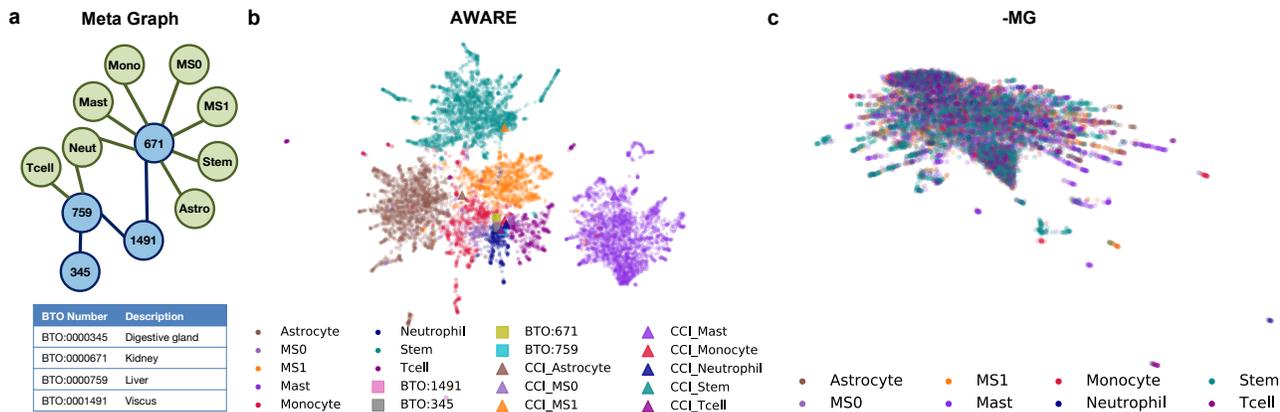


Figure 2. Visualization of contextual embeddings by AWARE and alternative approaches. Shown are 2D UMAP projections of our protein (circle), cell type (triangle), and tissue (square) embeddings from the (b) AWARE and (c) -MG methods. Unlike -MP (and GLOBAL, not shown due to space constraints), AWARE (and -PROTO, also not shown) clusters according to the cellular and tissue hierarchies imposed by the (a) meta graph. Notably, cell type embeddings (triangle) are proximal to their corresponding clusters of protein embeddings (circle). BTO, BRENDA Tissue Ontology; CCI, cell-cell interaction; MS0, Mesenchymal Stem Cell 0; MS1, Mesenchymal Stem Cell 1.

Table 1. Performance of protein embeddings on downstream prediction of disease-gene associations in 3 cell types. Reported are average Micro-F1 scores at k nearest neighbors, together with standard errors across 10 independent runs. Remarkably, as the number of neighbors k increases, GLOBAL and -MG’s performance decrease, likely due to poor separation of cell type specific protein embeddings.

METHOD	$k = 1$	$k = 5$	$k = 10$
GLOBAL	$0.24 \pm <0.01$	$0.20 \pm <0.01$	$0.06 \pm <0.01$
-MG	$0.15 \pm <0.01$	$0.13 \pm <0.01$	$0.02 \pm <0.01$
-PROTO	$0.27 \pm <0.01$	$0.28 \pm <0.01$	$0.11 \pm <0.01$
AWARE	$0.27 \pm <0.01$	$0.23 \pm <0.01$	$0.12 \pm <0.01$

Table 2. Evaluation of protein embeddings on downstream prediction task. Reported are average Recall@20 values and standard errors across 10 independent runs. A protein can be associated with 0, 1, or more diseases, with each association existing in the context of a specific cell type. The task is to predict cell type specific disease-gene associations from the learned protein embeddings. For instance, according to row #1, a Recall@20 value of 0.8 means that 80% of the genes truly associated with breast cancer when expressed in T cells are among the top 20 predicted by AWARE. Note that AWARE has slightly lower performance than -PROTO and -MG in two labels, likely due to the maximization of inter-class variation in AWARE, which can potentially separate similar proteins too much for the KNN to detect.

DISEASE	CELL TYPE	N_{TOTAL}	GLOBAL	-MG	-PROTO	AWARE
BREAST CANCER	T CELL	24	0.60 ± 0.16	0.90 ± 0.10	0.90 ± 0.10	0.80 ± 0.13
	ASTROCYTE	2	0.20 ± 0.13	0.20 ± 0.13	0.20 ± 0.13	0.20 ± 0.13
LIVER CANCER	T CELL	82	0.80 ± 0.13	0.50 ± 0.17	0.50 ± 0.17	0.50 ± 0.17
	ASTROCYTE	7	0.20 ± 0.13	0.30 ± 0.15	0.30 ± 0.15	0.30 ± 0.15
LUNG CANCER	T CELL	117	0.90 ± 0.10	0.60 ± 0.16	0.60 ± 0.16	0.60 ± 0.16
	ASTROCYTE	8	0.40 ± 0.16	0.40 ± 0.16	0.40 ± 0.16	0.40 ± 0.16
ALZHEIMER’S DISEASE (AD)	T CELL	37	0.50 ± 0.17	0.10 ± 0.10	$0.00 \pm <0.01$	0.20 ± 0.13
	ASTROCYTE	317	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$
	MONOCYTE	10	0.10 ± 0.10	0.20 ± 0.13	0.20 ± 0.13	0.20 ± 0.13
RHEUMATOID ARTHRITIS (RA)	T CELL	105	$1.00 \pm <0.01$	0.80 ± 0.13	0.90 ± 0.10	0.90 ± 0.10
	ASTROCYTE	15	0.10 ± 0.10	0.30 ± 0.15	0.30 ± 0.15	0.30 ± 0.15
	MONOCYTE	71	0.20 ± 0.13	0.60 ± 0.16	0.60 ± 0.16	0.60 ± 0.16
AUTISM SPECTRUM DISORDER (ASD)	T CELL	12	0.40 ± 0.16	0.40 ± 0.16	0.10 ± 0.10	0.10 ± 0.10
	ASTROCYTE	206	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$	0.90 ± 0.10
PANCREATIC DUCTAL ADENOCARCINOMA	T CELL	30	0.30 ± 0.15	0.80 ± 0.13	0.70 ± 0.15	0.60 ± 0.16
MULTIPLE SCLEROSIS (MS)	T CELL	90	0.60 ± 0.16	0.70 ± 0.15	0.50 ± 0.17	0.70 ± 0.15
	ASTROCYTE	239	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$
	MONOCYTE	4	0.10 ± 0.10	0.20 ± 0.13	0.20 ± 0.13	0.20 ± 0.13
ATHEROSCLEROSIS (ATH.)	T CELL	544	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$	$1.00 \pm <0.01$
	ASTROCYTE	23	0.40 ± 0.16	0.60 ± 0.16	0.60 ± 0.16	0.60 ± 0.16

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